

# Package: **BarcodingR** (via r-universe)

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barcodes.eval	<i>Barcodes Evaluation</i>
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---

## Description

Evaluate two barcodes using species identification success rate criteria.

## Usage

```
barcodes.eval(barcode1, barcode2, kmer1 = kmer1, kmer2 = kmer2)
```

## Arguments

barcode1	object of class "DNABin" based on barcode1, which contains taxon information.
barcode2	object of class "DNABin" based on barcode2, which contains taxon information.
kmer1	a numeric to indicate the length of kmer1 for barcode1, the optimal kmer could be found by the function optimize.kmer() before running this function.
kmer2	a numeric to indicate the length of kmer2 for barcode2, see above.

## Value

a list containing p\_value of prop.test(), and so on.

## Author(s)

Ai-bing ZHANG, PhD. CNU, Beijing, CHINA.

## References

zhangab2008 (at) mail. cnu. edu. cn.

**See Also**

prop.test()

**Examples**

```
data(TibetanMoth)
barcode1<-as.DNAbin(as.character(TibetanMoth[1:30,]))
barcode2<-barcode1
b.eval<-barcodes.eval(barcode1,barcode2,kmer1=1,kmer2=3)
b.eval
```

---

barcoding.gap

*Barcoding Gap Calculation*

---

**Description**

Calculation of DNA barcoding gap. Besides K2P distance, raw distance and euclidean could also be used for calculation DNA barcoding gap.

**Usage**

```
barcoding.gap(ref, dist = dist)
```

**Arguments**

ref	object of class "DNAbin" used as a reference dataset, which contains taxon information.
dist	a character string which takes one of ("raw","K80","euclidean").

**Value**

a list indicates the summary statistics of interspecific and intraspecific genetic distance, such as k2P distance.

**Note**

the current version of the function can only be used for protein-coding barcodes, such as, COI. The future version may incorporate calculation for non-coding barcodes, for instance, ITS1, ITS2.

**Author(s)**

Ai-bing ZHANG, PhD. CNU, Beijing, CHINA, contact at zhangab2008(at)mail.cnu.edu.cn

## References

Meyer, Christopher P., and Gustav Paulay. (2005). "DNA barcoding: error rates based on comprehensive sampling." *PLoS biology* 3.12: e422.

F.Jiang, Q. Jin, L. Liang, A.B. Zhang, and Z.H. Li.(2014). Existence of Species Complex Largely Reduced Barcoding Success for Invasive Species of Tephritidae: A Case Study in *Bactrocera* spp. *Mol Ecol Resour.* 14(6):1114-1128 DOI: 10.1111/1755-0998.12259.

## Examples

```
data(TibetanMoth)
TibetanMoth<-as.DNABin(as.character(TibetanMoth[1:20,]))
b.gap<-barcoding.gap(ref=TibetanMoth,dist="K80")
b.gap
```

---

barcoding.spe.identify

*Species Identification using Protein-coding Barcodes*

---

## Description

Species identification using protein-coding barcodes with different methods, including BP-based method (Zhang et al. 2008), fuzzy-set based method (Zhang et al. 2012), Bayesian-based method (Jin et al. 2013).

## Usage

```
barcoding.spe.identify(ref, que, method = "bpNewTraining")
```

## Arguments

ref	object of class "DNABin" used as a reference dataset, which contains taxon information.
que	object of class "DNABin", whose identities (species names) need to be inferred.
method	a character string indicating which method will be used to train model and/or infer species membership. One of these methods ("fuzzyId", "bpNewTraining", "bpNewTrainingOnly", "bpUseTrained", "Bayesian") should be specified.

## Value

a list containing model parameters used, species identification success rates using references, query sequences, species inferred, and corresponding confidence levels (bp probability for BP-based method / FMF values for fuzzy set theory based method / posterior probability for Bayesian method) when available.

**Note**

functions `fasta2DNABin()` from package:adegenet and `read.dna()` from package:ape were used to obtain DNABin object in our package. The former is used to read large aligned coding DNA barcodes, the latter unaligned ones. `ref` and `que` should be aligned with identical sequence length. We provided a pipeline to perform fast sequences alignment for reference and query sequences. Windows users could contact `zhangab2008(at)mail.cnu.edu.cn` for an exec version of the package. For very large DNA dataset, `read.fas()` package:phyloch is strongly suggested instead of `fasta2DNABin()` since the latter is very slow.

**Author(s)**

Ai-bing ZHANG, PhD. CNU, Beijing, CHINA. `zhangab2008(at)mail.cnu.edu.cn`

**References**

- Zhang, A. B., M. D. Hao, C. Q. Yang, and Z. Y. Shi. (2017). BarcodingR: an integrated R package for species identification using DNA barcodes. *Methods Ecol Evol.* 8(5):627-634. <https://besjournals.onlinelibrary.wiley.com/doi/10.1111/2017.12682>.
- Jin, Q., H.L. Han, X.M. Hu, X.H. Li, C.D. Zhu, S. Y. W. Ho, R. D. Ward, A.B. Zhang . (2013). Quantifying Species Diversity with a DNA Barcoding-Based Method: Tibetan Moth Species (Noctuidae) on the Qinghai-Tibetan Plateau. *PLoS One* 8: e644. <https://journals.plos.org/plosone/article?id=10.1371/journal.pone.006442>
- Zhang, A. B., C. Muster, H.B. Liang, C.D. Zhu, R. Crozier, P. Wan, J. Feng, R. D. Ward.(2012). A fuzzy-set-theory-based approach to analyse species membership in DNA barcoding. *Molecular Ecology*, 21(8):1848-63. <https://onlinelibrary.wiley.com/doi/10.1111/j.1365-294X.2011.05235.x>
- Zhang, A. B., D. S. Sikes, C. Muster, S. Q. Li. (2008). Inferring Species Membership using DNA sequences with Back-propagation Neural Networks. *Systematic Biology*, 57(2):202-215. <https://besjournals.onlinelibrary.wiley.com/doi/10.1111/2041-210X.12682>

**Examples**

```
data(TibetanMoth)
ref<-as.DNABin(as.character(TibetanMoth[1:5,]))
que<-as.DNABin(as.character(TibetanMoth[50:55,]))
bsi<-barcoding.spe.identify(ref, que, method = "fuzzyId")
bsi
bsi<-barcoding.spe.identify(ref, que, method = "bpNewTraining")
bsi
bsi<-barcoding.spe.identify(ref, que, method = "Bayesian")
bsi
```

---

barcoding.spe.identify2

*Species Identification Based on Fuzzy-set Method and kmer*

---

**Description**

Species identification based on fuzzy-set method (Zhang et al. 2012) and kmer.

**Usage**

```
barcoding.spe.identify2(ref, que, kmer = kmer, optimization = TRUE)
```

**Arguments**

ref	object of class "DNABin" used as a reference dataset, which contains taxon information.
que	object of class "DNABin", whose identities (species names) need to be inferred.
kmer	a numeric to indicate the length of maximum kmer to try in the range of 1 to kmer in case of optimization = TRUE, otherwise, only a certain length of kmer is used.
optimization	a character string, indicating whether different length of kmer (up to kmer) will be used or just a specified length of kmer will be used.

**Value**

a list indicating the identified species.

**Note**

read.dna() from package ape was used to obtain DNABin object for unaligned non-coding barcodes.

**Author(s)**

Ai-bing ZHANG, Cai-qing YANG, Meng-di HAO, CNU, Beijing, CHINA, contact at zhangab2008 (at) mail. cnu. edu. cn.

**References**

Zhang, A. B., M. D. Hao, C. Q. Yang, and Z. Y. Shi. (2017). BarcodingR: an integrated R package for species identification using DNA barcodes. *Methods Ecol Evol.* 8(5):627-634. <https://besjournals.onlinelibrary.wiley.com/doi/10.1111/2041-210X.12682>.

Jin, Q., H.L. Han, X.M. Hu, X.H. Li, C.D. Zhu, S. Y. W. Ho, R. D. Ward, A.B. Zhang . (2013). Quantifying Species Diversity with a DNA Barcoding-Based Method: Tibetan Moth Species (Noctuidae) on the Qinghai-Tibetan Plateau. *PloS One* 8: e644. <https://journals.plos.org/plosone/article?id=10.1371/journal.pone.006442>

Zhang, A. B., C. Muster, H.B. Liang, C.D. Zhu, R. Crozier, P. Wan, J. Feng, R. D. Ward.(2012). A fuzzy-set-theory-based approach to analyse species membership in DNA barcoding. *Molecular Ecology*, 21(8):1848-63. <https://onlinelibrary.wiley.com/doi/10.1111/j.1365-294X.2011.05235.x>

Zhang, A. B., D. S. Sikes, C. Muster, S. Q. Li. (2008). Inferring Species Membership using DNA sequences with Back-propagation Neural Networks. *Systematic Biology*, 57(2):202-215. <https://besjournals.onlinelibrary.wiley.com/doi/10.1111/2041-210X.12682>

**Examples**

```
data(pineMothITS2)
ref<-pineMothITS2
que<-ref
spe.id<-barcoding.spe.identify2(ref,que, kmer = 1, optimization = FALSE)
```

spe.id

---

bbsik

*Bp Barcoding Species Identify using Kmer*

---

## Description

Species identification using BP-based method for both protein-coding barcodes, for instance, COI, and non-coding barcodes, such as, ITS, using kmer statistics.

## Usage

```
bbsik(ref, que, kmer = kmer, UseBuiltModel = FALSE, lr = 5e-05, maxit = 1e+06)
```

## Arguments

ref	object of class "DNABin" used as a reference dataset, which contains taxon information.
que	object of class "DNABin", which needs to be inferred.
kmer	a numeric indicating the length of kmer used.
UseBuiltModel	logic value to indicate whether a built model is used or not.
lr	parameter for weight decay. Default 5e-5.
maxit	maximum number of iterations. Default 1e+6.

## Value

a list containing model parameters used, species identification success rates using references, query sequences, species inferred, and corresponding confidence levels (bp probability for BP-based method).

## Author(s)

Ai-bing ZHANG, Meng-di HAO, Cai-qing YANG, CNU, Beijing, CHINA. zhangab2008 (at) mail.cnu. edu.cn

## References

Zhang, A. B., D. S. Sikes, C. Muster, S. Q. Li. (2008). Inferring Species Membership using DNA sequences with Back-propagation Neural Networks. *Systematic Biology*, 57(2):202-215. <https://academic.oup.com/sysbio/article/57/2/202/1622290>

**Examples**

```
data(TibetanMoth)
ref<-as.DNABin(as.character(TibetanMoth[1:50,]))
que<-as.DNABin(as.character(TibetanMoth[51:60,]))
out<-bbsik(ref, que, kmer = 1, UseBuiltModel = FALSE)
out
out$convergence
out$success.rates.ref

data(pineMothITS2)
ref<-pineMothITS2
que<-pineMothITS2
out<-bbsik(ref, que, kmer = 1, UseBuiltModel = FALSE)
out
out$convergence
out$success.rates.ref
```

---

char2NumVector

*Character to Integer Vector*

---

**Description**

Conversion from a character vector to an integer vector.

**Usage**

```
char2NumVector(c)
```

**Arguments**

c                    character vector.

**Value**

an integer vector.

**Author(s)**

Ai-bing ZHANG, PhD. CNU, Beijing, CHINA. zhangab2008 (at) mail. cnu. edu.cn.

**References**

zhangab2008 (at) mail. cnu. edu. cn.



**Examples**

```
c<-c("a", "a", "b")
num<-char2NumVector(c)
num
```

---

compare2delimitations *Comparison between two Delimitations*

---

**Description**

Comparison between two delimitations of a group of samples, for instance, traditionally morphological delimitation and molecular delimitation (MOTU).

**Usage**

```
compare2delimitations(deli1, deli2)
```

**Arguments**

deli1	a character array (vector),containing a set of, for example, morphological identification (species names), to compare with
deli2	a character array (vector),containing a set of, molecular delimitation (MOTU).

**Value**

a list containing the adjusted Rand index comparing the two partitions (a scalar). This index has zero expected value in the case of random partition, and it is bounded above by 1 in the case of perfect agreement between two partitions; the numbers of matches, splits,merges, and corresponding percentage.

**Note**

This is for the same set of samples with two partitions/delimitations.

**Author(s)**

Ai-bing ZHANG, PhD. CNU, Beijing, CHINA.

**References**

L. Hubert and P. Arabie (1985) Comparing Partitions, Journal of the Classification 2:193-218.

## Examples

```
deli1<-c(1,1,1,1,1,1)
deli2<-c(1,1,2,1,1,3)
out<-compare2delimitations(deli1,deli2)
out
```

---

consensus.identify      *Consensus Identification*

---

## Description

Make consensus for identifications from two or more methods, usually for a set of query sequences.

## Usage

```
consensus.identify(identifiedBy2orMore)
```

## Arguments

identifiedBy2orMore  
an object of class "data.frame", containing (queIDs, as rownames), identified-ByMethod1,identifiedByMethod2,and so on.

## Value

a data frame with consensus.identification, and corresponding votes.

## Note

Suitable for case where a set of queries were identified by more than two methods.

## Author(s)

Ai-bing ZHANG, PhD. CNU, Beijing, CHINA, contact at zhangab2008(at)mail.cnu.edu.cn

## Examples

```
queIDs<-c("q1", "q2", "q3")

bp<-c("sp1", "sp1", "sp1")
bpk<-c("sp1", "sp1", "sp2")
bayes<-c("sp2", "sp1", "sp3")
fuzzyID<-c("sp1", "sp1", "sp2")
identifiedBy2orMore<-data.frame(bp=bp, bpk=bpk, bayes=bayes, fuzzyID=fuzzyID)
rownames(identifiedBy2orMore)<-queIDs<-c("q1", "q2", "q3")
ccs<-consensus.identify(identifiedBy2orMore)
```

---

`digitize.DNA`*Digitize DNABin*

---

**Description**

Digitize an object of DNABin.

**Usage**

```
digitize.DNA(seqs)
```

**Arguments**

`seqs` an object of DNABin.

**Value**

a numeric matrix of DNA sequences digitized.

**Author(s)**

Ai-bing ZHANG, PhD. CNU, Beijing, CHINA.

**References**

zhangab2008(at)mail.cnu.edu.cn

**Examples**

```
data(TibetanMoth)
digitized.DNA<-digitize.DNA(seqs=TibetanMoth)
digitized.DNA
```

---

`DNABin2kmerFreqMatrix` *Calculation of Kmer Frequency Matrix from DNABin for Both Reference and Query Sequences*

---

**Description**

Calculation of kmer frequency matrices from DNABin for both reference and query sequences.

**Usage**

```
DNABin2kmerFreqMatrix(ref, que, kmer = kmer)
```

**Arguments**

ref	Object of class "DNABin" used as a reference dataset, which contains taxon information.
que	Object of class "DNABin", which needs to be inferred.
kmer	a numeric to indicate the length of kmer used.

**Value**

kmer frequency matrices for both ref and que sequences, but only based on kmers found in ref!!!  
new kmers in que will be ignored.

**Author(s)**

Ai-bing ZHANG, PhD. CNU, Beijing, CHINA. zhangab2008(at)mail.cnu.edu.cn

**References**

zhangab2008(at)mail.cnu.edu.cn

**Examples**

```
data(TibetanMoth)
ref<-as.DNABin(as.character(TibetanMoth[1:50,]))
que<-as.DNABin(as.character(TibetanMoth[51:60,]))
out<-DNABin2kmerFreqMatrix(ref, que, kmer=3)
out
```

---

 FMF

---

*Fuzzy Membership Function Value*


---

**Description**

Calculation fuzzy membership function value given a distance from query to a potential species, maximal intraspecific variation of the potential species  $\theta_1$ , and minimal interspecific distance (here, the distance between the potential species and its nearest neighbor  $\theta_2$ ) (fuzzy-set based method, Zhang et al. 2012), different definition of distances could also be used.

**Usage**

```
FMF(xtheta12)
```

**Arguments**

xtheta12	a numerical vector containing three elements, a distance from query to a potential species, maximal or sd of intraspecific variation of the potential species $\theta_1$ , minimal or mean interspecific distance.
----------	--

**Value**

a numeric between 0 and 1.

**Note**

different definitions of distances could also be used.

**Author(s)**

Ai-bing ZHANG, Zhi-yong SHI. CNU, Beijing, CHINA, contact at zhangab2008(at)mail.cnu.edu.cn

**References**

Zhang, A. B., C. Muster, H.B. Liang, C.D. Zhu, R. Crozier, P. Wan, J. Feng, R. D. Ward.(2012). A fuzzy-set-theory-based approach to analyse species membership in DNA barcoding. *Molecular Ecology*, 21(8):1848-63.

**Examples**

```
xtheta12<-c(0.6289163,0.1465522,0.6379375)
FMF.out<-FMF(xtheta12)
FMF.out
```

---

FMFtheta12

*Calculate Intraspecific and Interspecific Variation*

---

**Description**

Calculation intraspecific variation (sd) of the potential species theta1, and mean interspecific distance (here, the mean distance between the potential species and its nearest neighbor theta2) (fuzzy-set based method, slightly modified from Zhang et al. 2012). The calculation was done for all species in the reference dataset.

**Usage**

```
FMFtheta12(ref)
```

**Arguments**

ref                    object of class "DNAbin" used as a reference dataset, which contains taxon information.

**Value**

a data frame containing intraspecific (sd, theta1) and interspecific variation (mean) of all species, and their corresponding nearest neighbor (NN).

**Author(s)**

Ai-bing ZHANG, PhD. CNU, Beijing, CHINA, contact at zhangab2008 (at) mail.cnu.edu.cn.

**References**

Zhang, A. B., C. Muster, H.B. Liang, C.D. Zhu, R. Crozier, P. Wan, J. Feng, R. D. Ward.(2012). A fuzzy-set-theory-based approach to analyse species membership in DNA barcoding. *Molecular Ecology*, 21(8):1848-63.

**Examples**

```
data(TibetanMoth)
ref<-as.DNABin(as.character(TibetanMoth[1:50,]))
FMF.theta12<-FMFtheta12(ref)
FMF.theta12
```

---

NAMES

*Extracts Lables of Samples*

---

**Description**

Extract sequence names from different objects of DNABin, including generated from fasta2DNABin() (package:adegenet), and read.dna() (package:ape).

**Usage**

```
NAMES(seqs)
```

**Arguments**

seqs                    object of class "DNABin", generated from fasta2DNABin() (package:adegenet), and read.dna() (package:ape).

**Value**

a character string array/vector.

**Author(s)**

Ai-bing ZHANG, PhD. CNU, Beijing, CHINA.

**References**

zhangab2008(at)mail.cnu.edu.cn

**Examples**

```
data(TibetanMoth)
seqNames<-NAMES(TibetanMoth)
seqNames
```

---

optimize.kmer	<i>Optimize kmer Length</i>
---------------	-----------------------------

---

**Description**

Optimize kmer length by trying kmers which length is in the range from 1 to max.kmer. The optimal kmer will have maximal species identification success rate.

**Usage**

```
optimize.kmer(ref, max.kmer = max.kmer)
```

**Arguments**

ref	object of class "DNABin" used as a reference dataset, which contains taxon information.
max.kmer	a numeric to indicate the length of maximal kmer.

**Value**

a numeric indicating the optimal kmer in the range examined.

**Author(s)**

Ai-bing ZHANG, Cai-qing YANG, Meng-di HAO, CNU, Beijing, CHINA.

**References**

zhangab2008 (at) mail. cnu. edu. cn/zhangab2008 (at) gmail.com.

**Examples**

```
data(TibetanMoth)
ref<-TibetanMoth[1:10,]
optimal.kmer<-optimize.kmer(ref,max.kmer=5)
```

---

pineMothCOI

*pine Moth COI*

---

## Description

COI DNA barcodes of Pine Moth in China.

## Usage

```
data("pineMothCOI")
```

## Format

The format is: 'DNABin' raw [1:140, 1:652] a t a a ... - attr(\*, "dimnames")=List of 2 ..\$ : chr [1:140] "A111,Lasiocampidae\_Dendrolimus\_punctatus" "A22,Lasiocampidae\_Dendrolimus\_punctatus" "A23,Lasiocampidae\_Dendrolimus\_punctatus" "A27,Lasiocampidae\_Dendrolimus\_punctatus" ... ..\$ : NULL

## Details

COI DNA barcodes of Pine Moth (Lasiocampidae) in China.

## Source

<http://dx.plos.org/10.1371/journal.pone.0064428>.

## References

Dai Q-Y, Gao Q, Wu C-S, Chesters D, Zhu C-D, and A.B. Zhang\*. (2012) Phylogenetic Reconstruction and DNA Barcoding for Closely Related Pine Moth Species (Dendrolimus) in China with Multiple Gene Markers. PLoS ONE 7(4): e32544.

## Examples

```
data(pineMothCOI)
pineMothCOI
```



---

pineMothITS1

*pine Moth ITS1*

---

### Description

ITS1 sequences of seven closely related pine moths species sampled through China.

### Usage

```
data("pineMothITS1")
```

### Format

The format is: List of 69 \$ A43,Lasiocampidae\_Dendrolimus\_punctatus: raw [1:698] 48 48 48 28 ... \$ A54,Lasiocampidae\_Dendrolimus\_punctatus: raw [1:696] 48 18 28 18 ... - attr(\*, "class")= chr "DNABin"

### Details

ITS1 used as DNA barcodes for closely related rine moth species (Dendrolimus) in China.

### Source

<http://dx.plos.org/10.1371/journal.pone.0064428>.

### References

Dai Q-Y, Gao Q, Wu C-S, Chesters D, Zhu C-D, and A.B. Zhang\*. (2012) Phylogenetic Reconstruction and DNA Barcoding for Closely Related Pine Moth Species (Dendrolimus) in China with Multiple Gene Markers. PLoS ONE 7(4): e32544.

### Examples

```
data(pineMothITS1)
pineMothITS1
```

---

pineMothITS2

*pine Moth ITS2*

---

### Description

ITS2 sequences of seven closely related pine moths species sampled in China.

### Usage

```
data("pineMothITS2")
```

**Format**

The format is: List of 97 \$ A22,Lasiocampidae\_Dendrolimus\_punctatus: raw [1:568] 48 48 48 28 ... \$ A23,Lasiocampidae\_Dendrolimus\_punctatus: raw [1:574] 48 18 28 18 ... \$ A29,Lasiocampidae\_Dendrolimus\_punctatus: raw [1:569] 88 48 48 48 ... \$ A52,Lasiocampidae\_Dendrolimus\_punctatus: raw [1:570] 48 18 28 18 ... - attr(\*, "class")= chr "DNABin"

**Details**

ITS2 used as DNA barcodes for closely related rine moth species (Dendrolimus) in China.

**Source**

<http://dx.plos.org/10.1371/journal.pone.0064428>.

**References**

Dai Q-Y, Gao Q, Wu C-S, Chesters D, Zhu C-D, and A.B. Zhang\*. (2012) Phylogenetic Reconstruction and DNA Barcoding for Closely Related Pine Moth Species (Dendrolimus) in China with Multiple Gene Markers. PLoS ONE 7(4): e32544.

**Examples**

```
data(pineMothITS2)
pineMothITS2
```

---

sample.ref

*Sample Random Datasets from References (DNABin)*

---

**Description**

Randomly sample reference data at different levels of taxon.

**Usage**

```
sample.ref(ref, sample.porp = 0.5, sample.level = "full")
```

**Arguments**

ref	Object of class "DNABin" used as a reference dataset, which contains taxon information.
sample.porp	a numeric value between 0 and 1, indicating proportion of samples to draw at each level of taxon.
sample.level	a character string choosing from c("full","family","genus","species").

**Value**

a list containing the selected samples and the samples left, in DNABin format stored in a matrix or a list.

**Note**

the ref must contain information on taxonomy, in format like, ">LS0909030M, Noctuidae\_Himalaea\_unica", i.e., "seqID, family\_genus\_species", or ">LS0909030M, Himalaea\_unica"; in case there is only one sample/individual for a taxon level, this sample will be retained in ref.selected.

**Author(s)**

Ai-bing ZHANG, PhD. CNU, Beijing, CHINA.

**References**

zhangab2008(at)mail.cnu.edu.cn;

**Examples**

```
data(TibetanMoth)
data(pineMothITS2)
ref<-TibetanMoth
ref2<-pineMothITS2
out<-sample.ref(ref, sample.porp=0.5, sample.level="full")
out
out2<-sample.ref(ref2, sample.porp=0.5, sample.level="full")
out2
```

---

save.ids

*Save Identifications*

---

**Description**

Output identified results to an outfile in tempory directory (found by tempdir() function).

**Usage**

```
save.ids(outfile = "identified.txt", ids)
```

**Arguments**

outfile	character string to indicate outfile name.
ids	object of class "BarcodingR", which contains identified taxon information.

**Value**

no value returned, but an output file.

**Author(s)**

Ai-bing ZHANG, PhD. CNU, Beijing, CHINA.

## References

zhangab2008(at)mail.cnu.edu.cn

## See Also

barcoding.spe.identify()

## Examples

```
data(TibetanMoth)
ref<-as.DNABin(as.character(TibetanMoth[1:50,]))
que<-as.DNABin(as.character(TibetanMoth[50:60,]))
bsi<-barcoding.spe.identify(ref, que, method = "fuzzyId")
bsi
save.ids(outfile="identified.txt",bsi)
```

---

summarize.ref

*Summarize Reference Data*

---

## Description

Summarize taxon information, sequence statistics, barcodes numbers per species for reference dataset.

## Usage

```
summarize.ref(ref, taxonStat = TRUE, seqStat = TRUE, barcodeStat = TRUE)
```

## Arguments

ref	object of class "DNABin" used as a reference dataset, which contains taxon information, or just an array containing taxon information only.
taxonStat	logic value to indicate whether the item is calculated only when ref is an object of class "DNABin".
seqStat	logic value to indicate whether the item is calculated
barcodeStat	logic value to indicate whether the item is calculated

## Value

a list containing taxon statistics, sequence statistics, population parameters, barcoding statistics ()

## Author(s)

Ai-bing ZHANG, Meng-di HAO, CNU, Beijing, CHINA.

## References

zhangab2008(at)mail.cnu.edu.cn/zhangab2008(at)gmail.com.

**Examples**

```
data(TibetanMoth)
s.r<-summarize.ref(TibetanMoth,taxonStat=TRUE,seqStat=TRUE,barcodeStat=TRUE)
s.r
```

TDR2

*TDR2 Species Membership Value***Description**

To calculate TDR value for a set of queries and one potential species. Its value is in the range of [0,1], 0 indicates extremely weak species membership, values close 1 indicating strong species membership.

**Usage**

```
TDR2(oneSpe, que, boot, boot2)
```

**Arguments**

oneSpe	object of class "DNABin" which contains DNA sequences from one species
que	object of class "DNABin" which contains DNA sequences different samples
boot	a numeric value indicating times of resampling along sequence columns
boot2	a numeric value indicating times of resampling along sequence rows (different samples)

**Value**

a numeric vector represents TDR values for each query against the species

**Note**

oneSpe and que should be the same in sequence length, i.e., they should be aligned in prior. It's strongly recommended that oneSpe should have large enough sample size, e.g., 20.

**Author(s)**

Ai-bing ZHANG, PhD. CNU, Beijing, CHINA, contact at zhangab2008(at)mail.cnu.edu.cn

**References**

Jin Q, L.J.He, A.B. Zhang\* (2012). A Simple 2D Non-Parametric Resampling Statistical Approach to Assess Confidence in Species Identification in DNA Barcoding-An Alternative to Likelihood and Bayesian Approaches. PLoS ONE 7(12): e50831. doi:10.1371/journal.pone.0050831. <http://dx.plos.org/10.1371/journal.pone.0050831>.

**Examples**

```

data(TibetanMoth)
sampleSpeNames<-NAMES(TibetanMoth)
Spp<-gsub(".", "", sampleSpeNames)
oneSpe<-TibetanMoth[grep("Macdunnoughia_crassisigna", Spp, value = FALSE, fixed = TRUE),]
oneSpe<-as.DNABin(as.character(oneSpe[1:5,]))
que<-TibetanMoth[grep("Agrotis_justa", Spp, value = FALSE, fixed = TRUE),]
que2<-oneSpe[1:2,]
out<-TDR2(oneSpe,que, boot=10,boot2=10) ### true false identification

```

---

TibetanMoth

*Tibetan Moth*


---

**Description**

COI DNA barcodes of Tibetan Moth in China.

**Usage**

```
data("TibetanMoth")
```

**Format**

The format is: 'DNABin' raw [1:319, 1:630] a t a a ... - attr(\*, "dimnames")=List of 2 ..\$ : chr [1:319] "LS0909030M, Noctuidae\_Himalaea\_unica" "LZ0827026M, Noctuidae\_Amphipyra\_pyramidea" "ML0829010M, Noctuidae\_Auchmis\_saga" "BM0830055M, Noctuidae\_Auchmis\_saga" ... ..\$ : NULL

**Details**

COI DNA barcodes of Tibetan Moth (Noctuidae) in China.

**Source**

<http://dx.plos.org/10.1371/journal.pone.0064428>.

**References**

Q. Jin, H.L. Han, X.M. Hu, X.H. Li, C.D. Zhu, S. Y. W. Ho, R. D. Ward, A.B. Zhang\* . (2013). Quantifying Species Diversity with a DNA Barcoding-Based Method: Tibetan Moth Species (Noctuidae) on the Qinghai-Tibetan Plateau. PloS One 8: e644.

**Examples**

```

data(TibetanMoth)
TibetanMoth

```

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